

Metabolic Reflow as a Therapy for Ischemic Brain Injury

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Abstract Ischemic neuronal damage is a common feature of occlusive strokes, hemorrhagic strokes, and traumatic brain injury. In addition, ischemia can be an anticipated or unanticipated complication of a variety of surgical procedures. Most therapeutic strategies for managing ischemic injury seek to re-establish blood flow, suppress neural metabolism, and/or limit specific cellular injury cascades. An alternative therapeutic approach is to enhance the delivery of metabolic substrates to ischemic tissue. This strategy is typified by efforts to increase tissue oxygenation by elevating the levels of circulating oxygen. Our studies are examining a complementary approach in which the delivery of metabolic substrates is enhanced by facilitating the diffusion of oxygen and glucose from the vasculature into neural tissue during ischemia. This is achieved by increasing the diffusivity of small molecules in aqueous solutions, such as plasma and interstitial fluid. The carotenoid compound, trans-sodium crocetin (TSC) is capable of increasing oxygen and glucose diffusivity, and our studies demonstrate that TSC increases cerebral tissue oxygenation in the penumbra of a focal ischemic event. In addition, TSC treatment reduces the volume of cerebral infarction in rodent models of both permanent and temporary focal ischemia. This strategy of “metabolic reflow” thus blunts the metabolic challenge in partially-perfused tissue and reduces ischemic neural injury.

Keywords Stroke · Neuroprotection · Oxygen · Diffusion · Metabolic reflow

Introduction

Ischemic neural damage can occur as a result of most types of stroke and time is of the essence for managing the ischemic insult to affected cells. Consequently, a primary goal for any therapeutic regimen is to reestablish blood flow to the ischemic tissue. In the case of occlusive (i.e. embolic and thrombotic) strokes, disruption of the intravascular impediment to blood flow, typically utilizing tissue plasminogen activator, is the principal therapeutic modality. In the case of hemorrhagic strokes, the situation is more complex because of the intrinsic challenges to stabilizing the hemorrhagic event. Subarachnoid hemorrhages resulting from the rupture of aneurysms on large cerebral arteries can be stabilized by surgical clipping or intravascular coiling. In contrast, intracerebral hemorrhages are less amenable to treatment. The management of hemorrhagic strokes is further complicated by the effects of residual extravascular blood in the subarachnoid and intraparenchymal spaces. Finally, downstream compromise to the microvasculature plays a role in the ischemic challenge produced by both occlusive and hemorrhagic strokes. These smaller vessels, although sometimes not directly impacted by an occlusion or hemorrhage are a key emerging target for optimizing blood supply after stroke.

Differential diagnosis of hemorrhagic versus occlusive stroke is a standard feature of the emergent response to stroke in order to ascertain whether thrombolytic therapy is appropriate to reestablish blood flow. Although this diagnosis consumes crucial time during an ongoing stroke, it is essential to limit the complications of rebleeding and hemorrhagic transformation. Consequently, an important goal for current experimental and clinical studies is to identify therapeutic modalities that can be administered rapidly and safely irrespective of stroke type. The predominant experimental approach to this problem focuses the inhibition of critical injury cascades. Numerous candidate agents targeting a wide variety of injury mechanisms have been

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identified and have shown considerable promise in preclinical studies of stroke. Unfortunately, human studies directed toward the inhibition of specific mechanisms of neural injury have not proven successful in advanced stage clinical trials. The reasons for these failures are several, including inadequate preclinical testing [9, 10, 29, 35, 38, 44, 53], and underpowered or inappropriately designed clinical trials [7, 10, 12, 15, 16, 26, 54]. Another plausible explanation for this lack of efficacy in late stage clinical trials is that drugs targeting a narrow range of injury mechanisms are incapable of overcoming the broad range of injury cascades that are set in motion by prolonged ischemia [60]. The future of neuroprotective therapy in the clinic may thus ultimately require a multimodal, poly-pharmacological approach to achieve success [41].

Two alternative approaches to limiting ischemic cellular injury involve either the suppression of overall metabolic demand [61] or the enhancement of metabolic supply [57]. The former approach seeks to broadly limit mechanisms of cellular injury by reducing metabolic demand and slowing the rates at which injury cascades can progress. The latter approach is also directed toward a broad inhibition of cellular injury, but does so by increasing metabolic supply to ischemic tissue and thus blunting the overall ischemic challenge. The studies described herein will focus on a novel form of the metabolic enhancement strategy using a pharmacologic intervention to increase the delivery of metabolic substrates to ischemic neural tissue.

Historically, therapeutic strategies to enhance the metabolic supply of ischemic tissue have generally utilized increased systemic oxygenation to improve tissue oxygenation. Hyperbaric and normobaric oxygenation have shown promise in limiting cellular injury and neurological function in animals models of stroke [4, 5, 11, 13, 18, 19, 24, 25, 28, 37, 39, 45, 47, 50, 51, 55–57, 59, 62]. In contrast, clinical trials have been more limited and have provided mixed outcomes to date [1–3, 6, 17, 32–34, 43, 48, 49, 63]. These mixed clinical outcomes can be ascribed in part to small sample sizes and relatively long delays prior to initiating treatment.

The underlying concept for the preceding oxygen enhancement studies is that the elevation of vascular oxygen raises the concentration gradient between blood and tissue and thus increases the movement of oxygen into the tissue compartment. However, it is important to note that the rate of movement of a small molecule, such as oxygen or glucose, is also dictated by its diffusion coefficient in the host medium. In the case of circulating blood, the plasma boundary layer is a key resistance for the movement of molecules from the vasculature into tissue [22, 23]. The diffusivity of oxygen and glucose in an aqueous solution, such as plasma, is dictated in part by the number of hydrogen bonds and intermolecular spacing among water molecules.

An increase in hydrogen bonding builds the “structure” of aqueous solutions and facilitates the diffusion of small molecules. It is thus possible to increase the access of vascular oxygen and glucose to tissue by modifying the diffusivity of these metabolic substrates. Trans-sodium crocetininate (TSC) is a carotenoid compound that has previously been shown to increase the diffusivity of small molecules, including oxygen and glucose, by facilitating structure building in aqueous media [27, 52]. Moreover, TSC has been shown to increase tissue oxygenation in multiple organs and to improve survival in a model of hemorrhagic shock [36, 42, 46]. The studies described herein examined the ability of TSC to increase cerebral oxygenation in areas of partial ischemia and its effects on neural injury in permanent and temporary models of focal brain ischemia [30].

Materials and Methods

All experimental protocols were approved by the University of Virginia Animal Care and Use Committee. Adult male Sprague-Dawley rats (330–370 g) underwent permanent or temporary focal ischemia [20] by clipping both common carotid arteries and the left middle cerebral artery (three-vessel occlusion: 3-VO). Using the permanent ischemia paradigm, TSC was administered at one of eight dosages, ranging from 0.023 to 4.580 mg/kg ($n = 7$ animals per group). Equivalent volumes of either Vehicle or TSC were injected into the femoral vein using a “bolus-infusion-bolus protocol”, as per the protocol of Okonkwo et al. [36]. Using this protocol, a bolus injection of 0.1 ml was administered 10 min after the onset of ischemia, followed by continuous infusion at 0.01 ml/min for 60 min. Thirty minutes after the end of infusion, another 0.1 ml bolus was injected. The dosages described here represent the total dosage of TSC administered using the bolus-infusion-bolus protocol. After 24 h of permanent ischemia, the animals were euthanized under deep anesthesia. The brains were sectioned coronally at a thickness of 2 mm and the sections were stained in 2% 2,3,5 triphenyltetrazolium chloride (TTC) in phosphate-buffered saline for 5 min at 37°C. Infarct size was measured and the total volume of infarction was corrected for swelling [30].

In the temporary model of focal ischemia, the 3-VO was maintained for 2 h after which the vessels were unclipped and reflow was established. These animals were euthanized at 22 h after establishing reflow. TSC was administered, as described above, beginning 10 min after the onset of ischemia. A dosage of 0.092 mg/kg was used based on the results of the dose-response study using the permanent ischemia model.

The effect of TSC on partial tissue oxygen levels (PtO₂) was also examined using the same temporary model of focal ischemia. A Licox probe was placed in the penumbra of the focal ischemic area in order to record tissue oxygenation. TSC was administered at a dosage of 0.92 mg/kg using the bolus-infusion-bolus protocol with the first bolus administered 10 min after the onset of ischemia. The levels of PtO₂ were recorded prior to ischemia for at least 20 min to obtain a stable, normoxic baseline. All recorded values were then normalized to this pre-ischemic baseline.

Results

Effect of TSC on Cerebral Infarction After Permanent Focal Ischemia

Treatment with TSC produced a dose-dependent reduction in cerebral injury. The dose-response curve was U-shaped with dosages ranging from 0.023 to 0.229 mg/kg producing significant reductions in infarct volume [30]. The most effective dosage, 0.092 mg/kg, reduced infarct volume by 58%.

Effect of TSC on Cerebral Infarction After Temporary Focal Ischemia

The optimal dosage of TSC for producing neuroprotection in the preceding dose-response experiment was tested for its effect on cerebral infarction produced by temporary (2 h) focal ischemia followed by 22 h of reperfusion. TSC treatment at a dosage of 0.092 mg/kg reduced infarct volume by 45% in this model of ischemia-reperfusion [30].

Effect of TSC on Oxygenation in the Ischemic Penumbra

Partial tissue oxygen levels in the ischemic penumbra of the 3-VO model were reduced by an average of 40–45% from baseline when ischemia was initiated [30]. Ten minutes after the onset of ischemia, animals were treated with either TSC or saline. In the TSC-treated animals, tissue oxygenation began to increase within approximately 10 min of the initial bolus injection. By the end of the second hour of ischemia PtO₂ in the penumbra of saline-treated animals was reduced by an average of 41% below baseline, while oxygen levels had recovered to only 20% below baseline in

the TSC-treated animals [30]. Upon reperfusion, tissue oxygenation increased well above baseline in both Vehicle-treated and TSC-treated animals. However, tissue hyperoxygenation during reperfusion was significantly less pronounced in TSC-treated animals than in Vehicle-treated animals.

Discussion

During an occlusive stroke, the ischemic penumbra is an area of partial blood flow that contains tissue at risk of being damaged and possibly recruited into cerebral infarction. The penumbra is generally viewed as a region that can be salvaged if vascular reperfusion is achieved within an adequate time frame and/or appropriate measures are taken to protect the tissue. The key determinants of how well penumbral tissue will withstand an ischemic challenge are the duration and depth of the ischemic event. Consequently, when it is feasible, a centerpiece of stroke management includes efforts to restore blood flow in order to limit the duration of metabolic challenge. Thrombolytic therapy can produce complete or partial recanalization with delays to reflow ranging from minutes to several hours after treatment [8, 40]. The delay to recanalization is compounded by the delay to receiving treatment, which typically is on the order of a couple of hours. Therefore, it is not unusual for stroke patients to experience ischemic events that persist for several hours. Although not always effective in establishing reflow, thrombolytic therapy remains the principal medical means for recanalization and a valuable strategy for curtailing the duration of an occlusive stroke.

The depth of an ischemic event can vary widely and the intensity of this challenge dictates the involvement and time course of various injury mechanisms [31]. Our current studies are examining a therapeutic approach designed to blunt the impact of partial ischemia by enhancing metabolic supply to at-risk tissue. Thus, this approach functionally attenuates the depth of ischemic challenge to the penumbra. Trans-sodium crocetininate was shown to substantially and significantly enhance tissue oxygenation in the penumbra of ongoing focal ischemia in the brain [30]. Moreover, TSC reduced neural damage in both permanent and temporary models of focal cerebral ischemia [30]. The protective actions of TSC are predicated on the concept of metabolic reflow, in which ischemic damage is attenuated by facilitating the delivery of metabolic substrates to at-risk tissue. The enhancement of tissue oxygenation is thought to result from the ability of TSC to increase the diffusivity of oxygen [52]. Nonetheless, in studies of this type, it is important to consider alternative explanations for the protective actions of any candidate therapy. Relevant to this issue are previous studies

examining the effects of TSC's structurally-similar parent compound, crocetin. These studies showed that crocetin did not exert significant effects on oxyhemoglobin saturation, oxygen solubility in blood, or blood flow [14, 21]. An increase in the diffusivity of metabolic substrates thus remains a parsimonious explanation for the observed increases in tissue oxygen and the resultant neural protection.

As discussed earlier, a key goal for ongoing and future studies will be to develop therapeutic modalities that can be implemented rapidly and safely irrespective of the type of stroke being treated. In this regard, it is noteworthy that preliminary data from our laboratory indicate that TSC also produces beneficial outcomes in an experimental model of intracranial hemorrhage [58]. It is therefore plausible that metabolic reflow therapy could be effective in treating both occlusive and hemorrhagic strokes. If so, this would obviate the need for a differential diagnosis of stroke type prior to initiating TSC treatment. Another key issue regarding the potential utility of metabolic reflow therapy concerns the time frame over which TSC can be effective. The current studies utilized a protocol in which TSC treatment was initiated soon after the onset of ischemia, which is not practicable in the context of most strokes. Ongoing studies are therefore examining the therapeutic window for TSC treatment in experimental models of stroke to ascertain whether delayed treatment is also effective in limiting ischemic damage.

In summary, our current studies have begun to characterize a novel therapeutic approach for treating ischemic neural injury. Metabolic reflow produced by TSC treatment can enhance the supply of metabolic substrates to at-risk tissue during ongoing partial ischemia. In addition, TSC treatment reduces cerebral infarction associated with permanent and temporary focal ischemia [30]. Future studies will be directed toward defining the utility of this strategy as an early intervention for the treatment of stroke, irrespective of the type of stroke that is occurring.

Conflict of interest statement We declare that we have no conflict of interest.

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References

- Al-Waili NS, Butler GJ, Beale J, Abdullah MS, Hamilton RW, Lee BY, et al. Hyperbaric oxygen in the treatment of patients with cerebral stroke, brain trauma, and neurologic disease. *Adv Ther*. 2005;22:659–678.
- Anderson DC, Bottini AG, Jagiella WM, Westphal B, Ford S, Rockswold GL, et al. A pilot study of hyperbaric oxygen in the treatment of human stroke. *Stroke* 1991;22:1137–1142.
- Bennett MH, Wasiaik J, Schnabel A, Kranke P, French C. Hyperbaric oxygen therapy for acute ischaemic stroke. *Cochrane Database Syst Rev*. 2005;20:CD004954
- Beynon C, Sun L, Marti HH, Heiland S, Veltkamp R. Delayed hyperbaric oxygenation is more effective than early prolonged normobaric hyperoxia in experimental focal cerebral ischemia. *Neurosci Lett*. 2007;425:141–145.
- Burt JT, Kapp JP, Smith RR. Hyperbaric oxygen and cerebral infarction in the gerbil. *Surg Neurol*. 1987;28:265–268.
- Carson S, McDonagh M, Russman B, Helfand M. Hyperbaric oxygen therapy for stroke: a systematic review of the evidence. *Clin Rehabil*. 2005;19:819–833.
- Cheng YD, Al-Khoury L, Zivin JA. Neuroprotection for ischemic stroke: two decades of success and failure. *NeuroRx* 2004;1:36–45.
- Delgado-Mederos R, Rovira A, Alvarez-Sabin J, Ribo M, Munuera J, Rubiera M, et al. Speed of tPA-induced clot lysis predicts DWI lesion evolution in acute stroke. *Stroke* 2007;38:955–960
- Dirnagl U. Bench to bedside: the quest for quality in experimental stroke research. *J Cereb Blood Flow Metab*. 2006;26:1465–1478.
- Donnan GA. The 2007 Feinberg lecture: a new road map for neuroprotection. *Stroke* 2008;39:242
- Eschenfelder CC, Krug R, Yusofi AF, Meyne JK, Herdegen T, Koch A, et al. Neuroprotection by oxygen in acute transient focal cerebral ischemia is dose dependent and shows superiority of hyperbaric oxygenation. *Cerebrovasc Dis*. 2008;25:193–201.
- Fisher M. Recommendations for advancing development of acute stroke therapies: stroke Therapy Academic Industry Roundtable 3. *Stroke* 2003;34:1539–1546.
- Flynn EP, Auer RN. Eubalic hyperoxemia and experimental cerebral infarction. *Ann Neurol*. 2002;52:566–572.
- Gainer JL, Rudolph DB, Caraway DL. The effect of crocetin on hemorrhagic shock in rats. *Circ Shock*. 1993;41:1–7.
- Ginsberg MD. Neuroprotection for ischemic stroke: past, present and future. *Neuropharmacology* 2008;55:363–389.
- Ginsberg MD. Current status of neuroprotection for cerebral ischemia: synoptic overview. *Stroke* 2009;40:S111–S114.
- Helms AK, Whelan HT, Torbey MT. Hyperbaric oxygen therapy of acute ischemic stroke. *Stroke*. 2007;38:1137; author reply 1138–1139.
- Henninger N, Bouley J, Nelligan JM, Sicard KM, Fisher M. Normobaric hyperoxia delays perfusion/diffusion mismatch evolution, reduces infarct volume, and differentially affects neuronal cell death pathways after suture middle cerebral artery occlusion in rats. *J Cereb Blood Flow Metab*. 2007;27:1632–1642.
- Henninger N, Kuppers-Tiedt L, Sicard KM, Gunther A, Schneider D, Schwab S. Neuroprotective effect of hyperbaric oxygen therapy monitored by MR-imaging after embolic stroke in rats. *Exp Neurol*. 2006;201:316–323.
- Hiramatsu K, Kassell NF, Goto Y, Soleau S, Lee KS. A reproducible model of reversible, focal, neocortical ischemia in Sprague-Dawley rat. *Acta Neurochir (Wien)*. 1993;120:66–71.
- Holloway GM, Gainer JL. The carotenoid crocetin enhances pulmonary oxygenation. *J Appl Physiol*. 1998;65:683–686.
- Huxley VH, Kutchai H. The effect of the red cell membrane and a diffusion boundary layer on the rate of oxygen uptake by human erythrocytes. *J Physiol*. 1981;316:75–83.
- Huxley VH, Kutchai H. Effect of diffusion boundary layers on the initial uptake of O₂ by red cells. Theory versus experiment. *Microvasc Res*. 1983;26:89–107.
- Kawamura S, Yasui N, Shirasawa M, Fukasawa H. Therapeutic effects of hyperbaric oxygenation on acute focal cerebral ischemia in rats. *Surg Neurol*. 1990;34:101–106.
- Kim HY, Singhal AB, Lo EH. Normobaric hyperoxia extends the reperfusion window in focal cerebral ischemia. *Ann Neurol*. 2005;57:571–575.
- Labiche LA, Grotta JC. Clinical trials for cytoprotection in stroke. *NeuroRx* 2004;1:46–70.
- Laidig K, Dagget V, Gainer J. Altering diffusivity in biological solutions via change of solution structure and dynamics. *J Am Chem Soc*. 1998;120:9394–9395.

28. Liu W, Sood R, Chen Q, Sakoglu U, Hendren J, Cetin O, et al. Normobaric hyperoxia inhibits NADPH oxidase-mediated matrix metalloproteinase-9 induction in cerebral microvessels in experimental stroke. *J Neurochem*. 2008;107:1196–1205.
29. Macleod MR, van der Worp HB, Sena ES, Howells DW, Dirnagl U, Donnan GA. Evidence for the efficacy of NXY-059 in experimental focal cerebral ischaemia is confounded by study quality. *Stroke* 2008;39:2824–2829.
30. Manabe H, Okonkwo DO, Gainer JL, Clarke R, Lee KS. Protection against focal ischemic injury to the brain by trans-sodium crocetin. *Journal of Neurosurgery* 2010;113(4). Epub:12/09 with podcast.
31. Moustafa RR, Baron J. Perfusion thresholds in cerebral ischemia. In: Donnan GA, Baron J, Davis SM, Sharp FR, editors. *The ischemic penumbra*. New York: Informa Healthcare USA, Inc.; 2002
32. Neubauer RA, End E. Hyperbaric oxygenation as an adjunct therapy in strokes due to thrombosis. A review of 122 patients. *Stroke* 1980;11:297–300.
33. Nighoghossian N, Trouillas P. Hyperbaric oxygen in the treatment of acute ischemic stroke: an unsettled issue. *J Neurol Sci*. 1997;150:27–31.
34. Nighoghossian N, Trouillas P, Adeleine P, Salord F. Hyperbaric oxygen in the treatment of acute ischemic stroke. A double-blind pilot study. *Stroke* 1995;26:1369–1372.
35. O'Collins VE, Macleod MR, Donnan GA, Horky LL, van der Worp BH, Howells DW. 1,026 experimental treatments in acute stroke. *Ann Neurol*. 2006;59:467–477.
36. Okonkwo DO, Wagner J, Melon DE, Alden T, Stone JR, Helm GA, et al. Trans-sodium crocetin increases oxygen delivery to brain parenchyma in rats on oxygen supplementation. *Neurosci Lett*. 2003;352:97–100.
37. Ostrowski RP, Colohan AR, Zhang JH. Mechanisms of hyperbaric oxygen-induced neuroprotection in a rat model of subarachnoid hemorrhage. *J Cereb Blood Flow Metab*. 2005;25:554–571.
38. Philip M, Benatar M, Fisher M, Savitz SI. Methodological quality of animal studies of neuroprotective agents currently in phase II/III acute ischemic stroke trials. *Stroke* 2009;40:577–581.
39. Qin Z, Karabiyikoglu M, Hua Y, Silbergleit R, He Y, Keep RF, et al. Hyperbaric oxygen-induced attenuation of hemorrhagic transformation after experimental focal transient cerebral ischemia. *Stroke* 2007;38:1362–1367.
40. Ribo M, Alvarez-Sabin J, Montaner J, Romero F, Delgado P, Rubiera M, et al. Temporal profile of recanalization after intravenous tissue plasminogen activator: selecting patients for rescue reperfusion techniques. *Stroke* 2006;37:1000–1004.
41. Rogalewski A, Schneider A, Ringelstein EB, Schabitz WR. Toward a multimodal neuroprotective treatment of stroke. *Stroke* 2006;37:1129–1136.
42. Roy JW, Graham MC, Griffin AM, Gainer JL. A novel fluid resuscitation therapy for hemorrhagic shock. *Shock* 1998;10:213–217.
43. Rusyniak DE, Kirk MA, May JD, Kao LW, Brizendine EJ, Welch JL, et al. Hyperbaric oxygen therapy in acute ischemic stroke: results of the hyperbaric oxygen in Acute Ischemic Stroke Trial Pilot Study. *Stroke* 2003;34:571–574.
44. Savitz SI. A critical appraisal of the NXY-059 neuroprotection studies for acute stroke: a need for more rigorous testing of neuroprotective agents in animal models of stroke. *Exp Neurol*. 2007;205:20–25.
45. Schabitz WR, Schade H, Heiland S, Kollmar R, Bardutzky J, Henninger N, et al. Neuroprotection by hyperbaric oxygenation after experimental focal cerebral ischemia monitored by MRI. *Stroke* 2004;35:1175–1179.
46. Seyde WC, McKernan DJ, Laudeman T, Gainer JL, Longnecker DE. Carotenoid compound crocetin improves cerebral oxygenation in hemorrhaged rats. *J Cereb Blood Flow Metab*. 1986;6:703–707.
47. Shin HK, Dunn AK, Jones PB, Boas DA, Lo EH, Moskowitz MA, et al. Normobaric hyperoxia improves cerebral blood flow and oxygenation, and inhibits peri-infarct depolarizations in experimental focal ischaemia. *Brain* 2007;130:1631–1642.
48. Singhal AB. A review of oxygen therapy in ischemic stroke. *Neurol Res*. 2007;29:173–183.
49. Singhal AB, Benner T, Roccatagliata L, Koroshetz WJ, Schaefer PW, Lo EH, et al. A pilot study of normobaric oxygen therapy in acute ischemic stroke. *Stroke* 2005;36:797–802.
50. Singhal AB, Dijkhuizen RM, Rosen BR, Lo EH. Normobaric hyperoxia reduces MRI diffusion abnormalities and infarct size in experimental stroke. *Neurology* 2002;58:945–952.
51. Singhal AB, Wang X, Sumii T, Mori T, Lo EH. Effects of normobaric hyperoxia in a rat model of focal cerebral ischemia-reperfusion. *J Cereb Blood Flow Metab*. 22:861–868.
52. Stennett AK, Dempsey GL, Gainer JL. trans-Sodium crocetin and diffusion enhancement. *J Phys Chem B*. 2006;110:18078–18080.
53. Stroke Therapy Academic Industry Roundtable. Recommendations for standards regarding preclinical neuroprotective and restorative drug development. *Stroke* 1999;30:2752–2758.
54. Stroke Therapy Academic Industry Roundtable. Recommendations for clinical trial evaluation of acute stroke therapies. *Stroke* 2001;32:1598–1606.
55. Sunami K, Takeda Y, Hashimoto M, Hirakawa M. Hyperbaric oxygen reduces infarct volume in rats by increasing oxygen supply to the ischemic periphery. *Crit Care Med*. 2000;28:2831–2836.
56. Veltkamp R, Siebing DA, Sun L, Heiland S, Bieber K, Marti HH, et al. Hyperbaric oxygen reduces blood-brain barrier damage and edema after transient focal cerebral ischemia. *Stroke* 2005;36:1679–1683.
57. Veltkamp R, Warner DS, Domoki F, Brinkhous AD, Toole JF, Busija DW. Hyperbaric oxygen decreases infarct size and behavioral deficit after transient focal cerebral ischemia in rats. *Brain Res*. 2000;853:68–73.
58. Wang Y, Yoshimura R, Manabe H, Lee KS. Effect of trans-sodium crocetin in a model of intracranial hemorrhage. *Society for Neuroscience Abstracts #472*. Washington, DC (2008).
59. Weinstein PR, Anderson GG, Telles DA. Results of hyperbaric oxygen therapy during temporary middle cerebral artery occlusion in unanesthetized cats. *Neurosurgery* 1987;20:518–524.
60. Yakovlev AG, Faden AI. Mechanisms of neural cell death: implications for development of neuroprotective treatment strategies. *NeuroRx* 2004;1:5–16.
61. Yenari M, Kitagawa K, Lyden P, Perez-Pinzon M. Metabolic downregulation: a key to successful neuroprotection? *Stroke* 2008;39:2910–2917.
62. Zhang JH, Lo T, Mychaskiw G, Colohan A. Mechanisms of hyperbaric oxygen and neuroprotection in stroke. *Pathophysiology* 2005;12:63–77.
63. Zhang JH, Singhal AB, Toole JF. Oxygen therapy in ischemic stroke. *Stroke* 2002;34:e152–3; author reply e153–e155.